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# Neocortex expansion is linked to size variations in gene families with chemotaxis, cell–cell signalling and immune response functions in mammals

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Increased brain size is thought to have played an important role in the evolution of mammals and is a highly variable trait across lineages. Variations in brain size are closely linked to corresponding variations in the size of the neocortex, a distinct mammalian evolutionary innovation. The genomic features that explain and/or accompany variations in the relative size of the neocortex remain unknown. By comparing the genomes of 28 mammalian species, we show that neocortical expansion relative to the rest of the brain is associated with variations in gene family size (GFS) of gene families that are significantly enriched in biological functions associated with chemotaxis, cell–cell signalling and immune response. Importantly, we find that previously reported GFS variations associated with increased brain size are largely accounted for by the stronger link between neocortex expansion and variations in the size of gene families. Moreover, genes within these families are more prominently expressed in the human neocortex during early compared with adult development. These results suggest that changes in GFS underlie morphological adaptations during brain evolution in mammalian lineages.

## 1. Introduction

Increased brain size in mammals when compared with other vertebrate taxa is thought to have played an important role in the expansion of this clade. Increased brain size during evolution has been previously related to increased behavioural complexity and the ability to cope with a changing environment [1,2]. However, the precise evolutionary drivers of brain size expansion in mammals and its relation to behavioural ability are still unclear and remain a topic of much interest and debate. This is complicated by the fact that different mammalian clades have differences in the degree of size-related changes in brain tissue [3]. Generally, large brains differ from small brains in having larger neuronal soma sizes [4], increased numbers of non-neuronal cells, in particular glia [5,6], and lower overall neuron density [7]. Large brains, however, are associated with a high metabolic cost [8–11] as well as higher demands of parental investment and delayed sexual maturation [12–16].

Brain size is a highly variable trait among mammalian and non-mammalian species with marked differences observed even between relatively close species [17–21]. Because brain size is closely associated with variations in body mass across species [22], comparative studies of brain size frequently use a corrected measure of brain size, known as encephalization index (Ei), which provides a

measure of how much brain size is above (or below) what is expected for a given body size. While  $E_i$  is commonly regarded as an index that aligns more closely with behavioural capacity [1,23,24], many studies have also related behavioural complexity directly to the actual size of specific brain regions as well as to relative brain size as a whole [25–29]. Changes in relative brain size (or  $E_i$ ) on the other hand, are not necessarily the result of a proportional expansion of all brain structures. In many mammalian lineages, most variations in encephalization index are closely linked to changes in the size of the neocortex [30–32], a distinctive structure of the mammalian brain and one of the most salient evolutionary innovations of the mammalian lineage [33–36].

The characteristic increase in the size of the neocortex relative to the rest of the brain has long been considered one of the primary targets of selection during mammalian brain evolution [37–39]. Increases in the absolute size of the neocortex are related to an increase in the number of functionally distinct neocortical areas [40–42], potentially allowing more complex information processing and the emergence of new behaviours [43]. In comparative studies in primates, for instance, relative size of the neocortex has been correlated with social group size [12,44,45] (but see [46]), and it has been speculated that the number of neocortical neurons may be a limiting factor in determining the number of social relationships mammals can effectively establish and manage [44]. More neocortical areas may be found in larger brains due to the lower marginal cost of devoting additional neural tissue to increasingly specialized functions [47], and an increasing number of neocortical areas may facilitate a more elaborate processing of sensory and motor information [48,49].

In the hominid lineage, the expansion of the neocortex is thought to have played a key role in the evolution of modern humans [50], including specialized areas involved in processing and production of language [51,52] as well as areas involved in identification of faces [53,54] and locations [55,56]. The neocortex in humans is widely regarded as the primary seat for the so-called higher cognitive functions, including self-awareness, consciousness, abstract reasoning and planning [57–63]. Development of the neocortex extends well into adolescence in humans and, although the structure of the layers in the neocortex is established during early prenatal development [64], the neocortex keeps growing in childhood and adolescence, reaching a peak in thickness on average at around 13 years of age, while myelination of some cortical regions can still continue after 20 years of age [65].

Despite the importance of the neocortex, the genomic features underlying its expansion during mammalian evolution remain poorly understood [30,66,67]. So far, there have been few efforts to identify features reflecting the genomic impact of brain evolution. Dorus and co-workers [68] reported accelerated sequence evolution of genes functioning in the nervous system during human origins, but this pattern was contested by later studies [69,70]. A genome-wide analysis of amino acid composition across 37 fully sequenced mammalian genomes showed that encephalization is significantly correlated with overall protein amino acid composition, although the causes of this pattern remain unclear [71].

Changes in gene family size (GFS) can reflect changes in the relative relevance of specific functions in an organism. Gene duplication events have been proposed to play a major role in the origin of novel gene functions and expression patterns [72,73]. Marked differences in GFS have been identified in

*Drosophila* and vertebrates, with families experiencing the largest changes being enriched in distinct biological functions [74–76]. Among mammals, marked differences in the number of olfactory receptors are likely to reflect variations in the reliance of different lineages on their sense of smell [77–80]. A recent study found that encephalization in mammalian lineages is associated with significant variations in GFS, with a significant enrichment of genes associated with immune system response, chemotaxis and cell–cell signalling functions among the most positively associated gene families [81]. Here, we investigate if variations in the relative size of the neocortex or neocortex to brain size ratio ( $N_r$ ) are associated with changes in GFS in mammalian lineages, and whether the extent to which any changes in GFS associated with  $N_r$  could explain previously reported associations between GFS variations and encephalization. We further explored whether any associated correlations between  $N_r$  and GFS are functionally reflected by the specific patterns of expression of  $N_r$ -associated families in the developing neocortex in humans.

## 2. Material and methods

### 2.1. Gene family annotations

Annotated gene families encompassing 28 fully sequenced mammalian genomes were obtained from Ensembl release 76 [82] (<http://www.ensembl.org>, Ensembl release 76). In the context of this annotation, Ensembl families are defined by clustering all Ensembl proteins along with metazoan sequences from UniProtKB. Any given gene family constitutes a group of related genes that includes both paralogues within the same species and orthologues and paralogues from other species. Any given gene can only be assigned to a single gene family. GFS in a given family for a given species was calculated as the total number of member genes contained in that gene family, for that particular species. In this study, we included all gene families with members present in at least six of the 28 mammalian species ( $n = 11\,943$ ). We excluded from this study any gene family with no variance in GFS across species.

### 2.2. Phenotype data

Body mass-corrected values of brain mass, known as encephalization index ( $E_i$ ), were computed as

$$E_i = \ln\left(\frac{\text{brain mass}}{\text{body mass}^b}\right).$$

The slope ( $b$ ) was estimated as 0.64 [83] based on a log–log least-squares linear regression of brain mass against body mass data from 493 mammalian species (table 1). Neocortex volumes were compiled from available literature (table 1), and include the grey and white matter of the cerebral cortex. Grey matter from palaeocortical structures (entorhinal cortex, schizocortex, hippocampus and amygdala) were excluded.  $N_r$  was defined as

$$N_r = \frac{\text{neocortex volume}}{\text{brain volume} - \text{neocortex volume}}$$

after Dunbar [44]. Maximum lifespan (MLSP) for each species was obtained from the animal ageing and longevity database, AnAge [100]. Brain region volumes and corresponding sources as well as encephalization indexes and MLSP for all included species are shown in table 1.

**Table 1.** Phenotypic traits for the 28 mammalian species analysed.

species name	common name	non-neocortex brain volume (cm <sup>3</sup> )	neocortex volume (cm <sup>3</sup> )	ref.	Nr	Ei	MLSP
<i>Ailuropoda melanoleuca</i>	giant panda	211.80935	136.43571	[84]	1.81	−2.014	36.8
<i>Callithrix jacchus</i>	marmoset	7.241	4.371	[85]	1.52	−1.627	16.5
<i>Canis familiaris</i>	dog (poodle)	458.273	177.753	[86]	0.63	−1.699	24
<i>Cavia porcellus</i>	guinea pig	4.671815	1.5798	[87]	0.51	−2.948	12
<i>Echinops telfairi</i>	lesser hedgehog tenrec	0.566	0.0515	[85]	0.1	−3.274	19
<i>Erinaceus europaeus</i>	hedgehog	3.05	0.522	[85]	0.21	−2.863	11.7
<i>Gorilla gorilla</i>	gorilla	470.359	341.444	[85]	2.65	−1.415	55.4
<i>Homo sapiens</i>	human	1251.847	1006.525	[85]	4.1	0.152	122.5
<i>Loxodonta africana</i>	elephant	3886.7	2460.1	[88]	1.72	−1.082	65
<i>Macaca mulatta</i>	macaque	87.896	63.482	[85]	2.6	−1.192	40
<i>Macropus eugenii</i>	wallaby	11.6637	4.3987	[89]	0.61	−2.207	15.1
<i>Microcebus murinus</i>	mouse lemur	1.68	0.74	[85]	0.79	−1.985	18.2
<i>Mus musculus</i>	mouse (C57BL/6J)	0.48	0.12	[90]	0.32	−2.832	4
<i>Mustela putorius furo</i>	European polecat	8.8996	4.147	[91]	0.87	−2.548	11.1
<i>Ornithorhynchus anatinus</i>	platypus	8.57145	4.09928	[92]	0.92	−2.219	22.6
<i>Ovis aries</i>	sheep	100.332	53.793	[93]	1.16	−1.961	22.8
<i>Pan troglodytes</i>	chimpanzee	382.103	291.592	[85]	3.22	−0.948	59.4
<i>Papio anubis</i>	olive baboon	190.957	140.142	[85]	2.76	−1.178	37.5
<i>Pongo abelii</i>	orangutan	304.2	219.8	[94]	2.6	−0.892	59
<i>Procavia capensis</i>	hyrax	12.68	5.54	[95]	0.78	−2.255	14.8
<i>Pteropus vampyrus</i>	megabat	8.89	3.61	[96]	0.68	−2.204	20.9
<i>Rattus norvegicus</i>	rat	1.69	0.58	[95]	0.52	−2.861	5
<i>Sarcophilus harrisii</i>	Tasmanian devil	15.1517	3.7334	[89]	0.33	−2.792	13
<i>Sorex araneus</i>	shrew	0.188	0.0264	[85]	0.16	−2.832	3.2
<i>Sus scrofa</i>	pig	106.660	54.3913	[97]	1.04	−2.468	27
<i>Tarsius syrichta</i>	tarsier	3.393	1.768	[85]	1.09	−1.795	16
<i>Tursiops truncatus</i>	dolphin	1376.976	1088.615	[98]	3.78	−0.321	51.6
<i>Vicugna pacos</i>	alpaca	181.467	101.81	[99]	1.28	−1.688	25.8

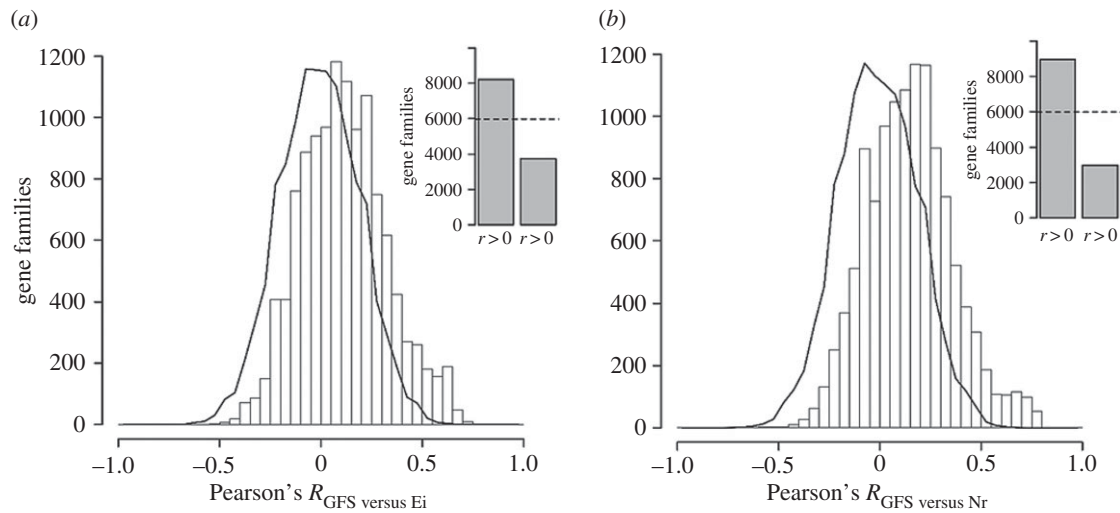
### 2.3. Correlation coefficients of gene family size and different phenotypes

Pearson's correlations between GFS values and the three phenotypes, Ei, Nr or MLSP, for all 11 943 gene families included in the study, were calculated using R-based statistical functions. To determine the statistical significance of any potential shift in the distribution of Pearson's correlation coefficients when compared to random expectation, 10 000 Monte Carlo simulations of the expected distribution based on random permutations of GFS values across species were conducted and contrasted with the observed distribution of correlation coefficients using a Z-score test.

### 2.4. Confounding variables and phylogenetically controlled correlations

In order to remove the effect of Ei and MLSP on Nr, we calculated residuals for the multivariate regression of  $Nr \sim Ei + MLSP$  (with Nr as the response variable and Ei and MLSP as

independent covariates). For consistency, we used the exact same approach to obtain similar corrected estimates for all GFS values after correcting for any potential effects of Ei and MLSP. This was done by extracting the residuals for the multivariate regression  $GFS \sim Ei + MLSP$  for each individual gene family. The resulting sets of residuals were then used to obtain phylogenetic independent contrasts (PIC) to further account for any effect of phylogenetic relationships on these variables [101]. The resulting independent contrasts were finally used to assess the final corrected association between Nr and GFS by simply using standard Pearson's correlations forced through the origin. The same analysis was carried but using Nr and MLSP instead as independent covariates to generate residuals for all Ei and GFS values, from the multivariate regressions  $Ei \sim Nr + MLSP$  and  $GFS \sim Nr + MLSP$  respectively, followed by extraction of the corresponding PIC to assess the unbiased association between Ei and GFS (figure 1). PIC analysis was computed using the ape package in R [102]. Ultrametric phylogeny of the 28 analysed mammalian species was obtained from TimeTree database [103] (<http://www.timetree.org/>. TimeTree2).



**Figure 1.** Enrichment of gene family size variations (GFS) in line with increased encephalization index (Ei) and neocortex to brain size ratio (Nr) in mammals. (a) Histogram showing the distribution of correlation coefficients for GFS and Ei in 11 943 gene families encompassing 28 mammalian genomes. (b) Histogram showing the distribution of correlation coefficients for GFS and Nr in 11 943 gene families encompassing 28 mammalian genomes. In each figure, an estimation of the expected distribution derived from 10 000 Monte Carlo simulations is represented by the solid line. Inset: distribution of positive and negative correlations relative to the expected distribution (dashed line).

## 2.5. Gene Ontology term enrichment

Gene Ontology (GO) annotations per species for biological process domains were obtained from Ensembl's Biomart release 76 [82]. A GO term was associated with a family whenever that term was linked to any of its members in any species. To minimize the effect of very small functional categories, only terms linked to at least 200 gene families were examined ( $n = 116$ ). GO terms with less than 200 gene families were assigned to a 'small biological process GO terms' category while gene families not annotated to any GO term in any species were grouped into a 'not annotated' category. Enrichment analysis of these GO terms was carried out as described in Castillo-Morales *et al.* [81]. Briefly, over-representation of genes associated with specific GO terms was assessed by counting the number of gene families assigned to each GO term within the analysed set of gene families. Statistical significance was numerically assessed by obtaining the expected number of families per GO in 1000 equally sized random samples derived from the overall population of gene families. Because genes vary in the number of GO terms associated with them, we adjusted for differences in the density of GO annotations between the test and background samples, by dividing the family counts per GO from each sample by the sample's average number of GO annotations per family.

## 2.6. Gene expression before and after full cortical maturation

RNAseq RPKM normalized expression data summarized to genes were obtained from the NIMH Transcriptional Atlas of Human Brain Development database [104] (<http://brain-span.org>. BrainSpan Atlas of the Developing Human Brain) for a total of 143 post-mortem human brain samples corresponding to 11 cortical regions across 13 different ages. The cortical regions include primary auditory cortex (core) (A1C), dorsolateral prefrontal cortex (DFC), posteroinferior (ventral) parietal cortex (IPC), inferolateral temporal cortex (area TEv, area 20) (ITC), primary motor cortex (area M1,

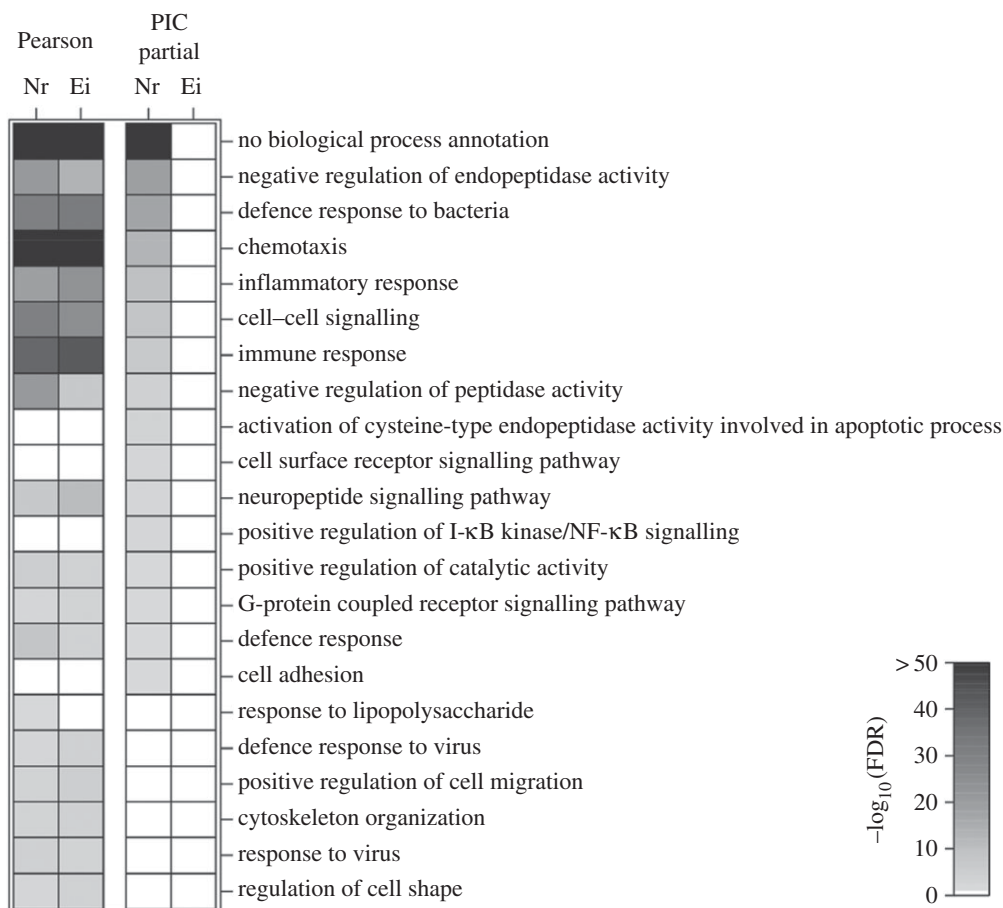
area 4) (M1C), anterior (rostral) cingulate (medial prefrontal) cortex (MFC), orbitofrontal cortex (OFC), primary somatosensory cortex (area S1, areas 3,1,2) (S1C), posterior (caudal) superior temporal cortex (area TAc) (STC), primary visual cortex (striate cortex, area V1/17) (V1C) and ventrolateral prefrontal cortex (VFC). The samples covered developmental stages 16, 24, 37 post-conception weeks, four months after birth and 1, 3, 8, 13, 19, 21, 30, 36 and 37 years old. Gene expression data were further normalized against the total expression per sample, and divided into two developmental groups, corresponding to the periods before and after full maturation of cortical thickness, which occurs at about 13 years of age in humans [65]. For each gene, expression levels were averaged across stages and structures of the same developmental window and comparisons between developmental windows were carried out by means of paired Wilcoxon tests.

## 3. Results

In order to assess the association between gene family size, GFS, and neocortex expansion, Nr, values were compiled from the literature for 28 mammalian species with fully sequenced genomes (table 1). GFS was calculated for a total of 11 943 non-overlapping families. Pearson's correlation coefficients between GFS and Nr were then calculated for each gene family. We found a significant over-representation of gene families with positive associations between GFS and Nr (figure 1) ( $\chi^2 = 2973.263083$ ,  $p < 1 \times 10^{-20}$ ). A Monte Carlo simulation showed that the observed shift in the distribution towards positive values is statistically significant when compared with random expectation ( $Z$ -score for observed mean  $R = 2.225819868$ ,  $p = 0.013$ ).

In order to assess whether the observed bias towards strong correlations between GFS and Nr preferentially involves gene families associated with specific biological functions (as opposed to random sets of gene families), we assessed the statistical over-representation of functional annotations (annotated GO terms per gene family, see Material and





**Figure 2.** Gene ontology enrichment analysis of families with gene family size (GFS) variations in line with encephalization index (Ei) and neocortex to brain size ratio (Nr). Heatmap of the significance of the over-representation of GO terms (expressed as Benjamini–Hochberg (BH)-corrected  $p$ -value) among gene families most significantly associated with Ei and Nr. First two columns correspond to gene families with the most significant association between GFS and Ei or Nr, respectively ( $r_{Nr, GFS} > 0$ ,  $FDR < 0.05$ ,  $n = 440$  and  $r_{Ei, GFS} > 0$ ,  $FDR < 0.05$ ,  $n = 321$ ). Third and fourth columns represent GO terms enriched among gene families whose GFS variations display the most significant association with one of the brain phenotypes after accounting for the shared variance with the other neural phenotype, as well as the phylogenetic relationship of the analysed species using independent contrast analysis ( $r_{PIC(Nr \sim Ei + MLSP), PIC(GFS \sim Ei + MLSP)} > 0$ ,  $FDR < 0.05$ ,  $n = 272$  and  $r_{PIC(Ei \sim Nr + MLSP), PIC(GFS \sim Nr + MLSP)} > 0$ ,  $FDR < 0.05$ ,  $n = 0$  respectively). Only GO terms significantly enriched after BH multiple testing correction are shown in the figures.

methods) for the 440 gene families found to be significantly associated with Nr ( $r_{Nr, GFS} > 0$  and  $FDR < 0.05$ ). A total of 18 GO functional categories were found to be significantly enriched ( $FDR < 0.05$ ) among Nr-associated gene families including immune response, negative regulation of endopeptidase activity, chemotaxis, cell–cell signalling, neuropeptide signalling pathway and G-coupled receptor signalling pathway (figure 2). Notably, genes with no functional annotations showed the highest over-representation.

As Nr is known to be highly correlated to relative brain size, the observed association between Nr and changes in GFS could be explained in principle by a previously reported association between GFS and relative brain size [81]. Indeed, after calculating correlation coefficients between GFS and Ei (a commonly used index of brain size relative to body mass) for each gene family in the same set of 28 species used in this study (figure 1), we also found a significant shift in the distribution favouring positive associations. This finding is consistent with a previously reported study using a larger set of 39 species [81]. The shift in the distribution of correlation coefficient values, however, was found to be stronger for Nr compared with Ei with the significance of the deviation for the latter being lower, ( $Z_{Ei} = 1.70943$ ,  $p = 0.044$ , figure 1). Functional annotation enrichment analysis revealed a total of 17 GO term categories enriched among the set of gene families found to be significantly associated with Ei ( $r_{Ei, GFS} > 0$  and

$FDR < 0.05$ ), with a strong overlap with the 18 GO functional categories found overrepresented among Nr-associated gene families (Jaccard index = 0.67) (figure 2).

To assess whether variations in GFS associated with Nr are secondary to the relationships between Ei and GFS, we obtained Ei-corrected residuals for Nr and GFS. In addition, due to a known relationship between encephalization and MLSP in mammals we also corrected for the potential effect of this trait [83,105]. Finally, in order to remove any phylogenetic signal from the correlations between our traits of interest and GFS arising from interrelatedness among species, we used the above Nr and GFS residuals to conduct a PIC analysis (see Material and methods). This phylogenetically corrected analysis of GFS and Nr residuals revealed a total of 272 families significantly associated with Nr after correction for multiple testing (phylogenetically controlled  $r$ 's  $> 0$ ,  $FDR < 0.05$ ; electronic supplementary material, table S1). By contrast, phylogenetically controlled correlations between equivalent GFS and Ei residuals (correcting for the effect of Nr and MLSP, see Material and methods) resulted in no gene families with a significant association after correcting for multiple testing. Nr-associated gene families after this correction against confounding variables were found to be enriched in GO terms including immune response, negative regulation of endopeptidase activity, chemotaxis, cell–cell signalling and neuropeptide signalling pathway (figure 2).

If the association between GFS and Nr responds to the functional demands imposed by the development of a large neocortex, we should expect genes associated with families displaying a high correlation with Nr to also display a pronounced level of activity prior to full cortical maturation (when full cortical thickness is reached), compared with later stages. To this end, we used available gene expression data derived from human neocortex obtained from the BrainSpan Atlas of the Developing Human Brain [104] (see Material and methods). We found that gene members of this set of families showed higher expression levels during human development prior to the neocortex reaching maximum thickness (which in humans occurs around the age of 13 years) compared with later stages, reflecting a transcriptional signature of the potential involvement of some of these genes in the development of the neocortex ( $p = 0.00013$ ).

## 4. Discussion

The expansion of the neocortex observed in several mammalian lineages is considered to be linked to a proliferation of new cortical areas driving increased cognitive capabilities [40,106]. The genomic drivers shaping the evolution of the brain and its morphology remain, however, poorly understood. By comparing the genomes of 28 mammalian species, here we have assessed the potential association between changes in GFS and the expansion of the neocortex. We show that neocortical expansion is indeed strongly and specifically associated with variations in GFS in mammals. Furthermore, variations in relative neocortical size account for a high proportion of the previously reported links between GFS and changes in encephalization across mammalian species [81]. This suggests that changes in GFS in line with relative brain size in mammals are actually secondary to the underlying correlation between neocortex size and encephalization. Analysis of available human neocortex gene expression data revealed that genes in families strongly and specifically associated with neocortex size variations also show significantly higher levels of expression at stages of development before (but not after) maximal cortical development is reached in humans, thereby supporting a functional role for these gene families in the ontological development of a large neocortex. Among the 272 gene families whose size was found robustly correlated with relative neocortex size, even after correcting for encephalization, MLSP and phylogenetic relationships, 16 distinct biological functions (GO terms) were found to be significantly overrepresented. Among these, cell–cell signalling and chemotaxis are known to play critical roles in the development and maintenance of the nervous system. Example of Nr-associated gene families annotated to these functions are the tyrosine kinase precursor family (ENSMF00730001521921), encoding receptor protein-tyrosine kinases and widely known to promote cell survival, proliferation, adhesion and migration in the central nervous system [107–109]. The leukotriene B4 receptor 2 family (ENSMF00680001303697) includes leukotriene B4, a proinflammatory signalling molecule which has been shown to mediate regulation of neural stem cell proliferation and differentiation [110].

Several immune-related biological functions (inflammatory response, defence response to bacteria, immune response, defence response and positive regulation of I- $\kappa$ B

kinase/NF- $\kappa$ B signalling) were also enriched among Nr-associated gene families. Along these lines in recent years, numerous immune-related signalling and regulatory components have also been shown to play key physiological roles in the developing and adult nervous system (for a review see [111]). This involvement of individual immune-related signalling components in neural functions has been shown to be part of a wider genetic network of immune-related molecules acting as an intrinsic component of the neural-specific regulatory machinery that ultimately shapes the normal development of the nervous system [112]. Thus, for instance, members of the tumour necrosis factor (TNF) receptor superfamily (ENSMF00500000273041, a gene family found to be highly associated with neocortex expansion here), are themselves part of the extensively studied canonical pathway of activation of the transcription factor NF- $\kappa$ B during early development of the nervous system [111].

Interestingly, gene families with no reported functional annotations for any of its members in any species showed the highest enrichment among the gene families with the highest positive associations with relative neocortex size. Among these families, we found the neuroblastoma breakpoint gene family (ENSMF00250000000879), whose members contain DUF1220 domains. DUF1220 domains have been previously linked to brain and cortical expansion in primate species, particularly in the human lineage [113,114]. Polymorphic deletions and duplications of DUF1220 domains have been associated with brain size variations in normal individuals from different human populations as well in pathological cases including microcephaly and macrocephaly [115,116]. Moreover, it has been proposed that proteins containing this domain have an important role during cortical neurogenesis, as they promote proliferation in neural stem cells [113], and during normal development they are expressed in the sub-ventricular zone precisely during the period of cortical neurogenesis [114].

Of particular importance to build a larger neocortex is the control of successive rounds of proliferation during early development, where the interplay between symmetric and asymmetric cell division is thought to be critical in shaping the particular morphology of the neocortex [117]. Consistent with this, one gene family with significant GFS changes in line with increased relative neocortex size is the ENSFM00250000003440 gene family of epithelial cell adhesion molecules, which in turn include known human developmental regulators such as EPCAM and TACSTD2. EPCAM has been shown to be involved in cell proliferation, differentiation and migration in diverse cell types [118,119] and could thus play an important role in neocortex development. A more numerous gene family found was the speedy gene family (ENSMF00740001589497), which encodes proteins able to bind CDKs but having no similarity with cyclins, and some of its members are known to play a role in the regulation of cell cycle [120,121]. While a great variation in gene numbers across species has been documented in this family [122], here we report the first evidence of a strong association between these variations in this family and relative neocortex size in mammals.

## 5. Conclusion

In summary, we have identified a set of gene families whose sizes are positively associated with an expanded neocortex,

providing new insights into neocortex evolution. Moreover, as aberrant development and degeneration of cortical neurons has been linked with a variety of mental health pathologies and dementias [123,124], identifying genomic signatures associated with the evolution of larger brain size and neocortex expansion will critically contribute to our understanding of the molecular pathways involved in the development and maintenance of cortical areas in highly encephalized mammals including humans. As these pathways may not be present or developed to the same extent in less encephalized mammalian species, our finding could help to fill existing gaps in current knowledge gained from widely used rodent models.

## References

- Deaner RO, Isler K, Burkart J, van Schaik C. 2007 Overall brain size, and not encephalization quotient, best predicts cognitive ability across non-human primates. *Brain Behav. Evol.* **70**, 115–124. (doi:10.1159/000102973)
- Reader SM, Laland KN. 2002 Social intelligence, innovation, and enhanced brain size in primates. *Proc. Natl Acad. Sci. USA* **99**, 4436–4441. (doi:10.1073/pnas.062041299)
- Herculano-Houzel S. 2012 Neuronal scaling rules for primate brains: the primate advantage. *Progr. Brain Res.* **195**, 325–340. (doi:10.1016/B978-0-444-53860-4.00015-5)
- Haug H. 1987 Brain sizes, surfaces, and neuronal sizes of the cortex cerebri: a stereological investigation of man and his variability and a comparison with some mammals (primates, whales, marsupials, insectivores, and one elephant). *Am. J. Anat.* **180**, 126–142. (doi:10.1002/aja.1001800203)
- Herculano-Houzel S. 2014 The glia/neuron ratio: how it varies uniformly across brain structures and species and what that means for brain physiology and evolution. *Glia* **62**, 1377–1391. (doi:10.1002/glia.22683)
- Lewitus E, Hof PR, Sherwood CC. 2012 Phylogenetic comparison of neuron and glia densities in the primary visual cortex and hippocampus of carnivores and primates. *Evol. Int. J. Org. Evol.* **66**, 2551–2563. (doi:10.1111/j.1558-5646.2012.01601.x)
- Tower DB. 1954 Structural and functional organization of mammalian cerebral cortex; the correlation of neurone density with brain size; cortical neurone density in the fin whale (*Balaenoptera physalus* L.) with a note on the cortical neurone density in the Indian elephant. *J. Comp. Neurol.* **101**, 19–51. (doi:10.1002/cne.901010103)
- Aiello LC, Wheeler P. 1995 The expensive-tissue hypothesis: the brain and the digestive system in human and primate evolution. *Curr. Anthropol.* **36**, 199–221. (doi:10.2307/2744104)
- Navarrete A, van Schaik CP, Isler K. 2011 Energetics and the evolution of human brain size. *Nature* **480**, 91–93. (doi:10.1038/nature10629)
- Fish JL, Lockwood CA. 2003 Dietary constraints on encephalization in primates. *Am. J. Phys. Anthropol.* **120**, 171–181. (doi:10.1002/ajpa.10136)
- Isler K, van Schaik CP. 2006 Metabolic costs of brain size evolution. *Biol. Lett.* **2**, 557–560. (doi:10.1098/rsbl.2006.0538)
- Barton RA, Capellini I. 2011 Maternal investment, life histories, and the costs of brain growth in mammals. *Proc. Natl Acad. Sci. USA* **108**, 6169–6174. (doi:10.1073/pnas.1019140108)
- Barrickman NL, Bastian ML, Isler K, van Schaik CP. 2008 Life history costs and benefits of encephalization: a comparative test using data from long-term studies of primates in the wild. *J. Hum. Evol.* **54**, 568–590. (doi:10.1016/j.jhevol.2007.08.012)
- Deaner RO, Barton RA, Van Schaik C. 2003 Primate brains and life histories: renewing the connection. In *Primates life histories and socioecology* (eds PM Kappeler, ME Pereira), pp. 233–265. Chicago, IL: The University of Chicago Press.
- Finarelli JA. 2010 Does encephalization correlate with life history or metabolic rate in Carnivora? *Biol. Lett.* **6**, 350–353. (doi:10.1098/rsbl.2009.0787)
- Isler K, van Schaik CP. 2009 The expensive brain: a framework for explaining evolutionary changes in brain size. *J. Hum. Evol.* **57**, 392–400. (doi:10.1016/j.jhevol.2009.04.009)
- Harvey PH, Clutton-Brock TH, Mace GM. 1980 Brain size and ecology in small mammals and primates. *Proc. Natl Acad. Sci. USA* **77**, 4387–4389. (doi:10.1073/pnas.77.7.4387)
- Huber R, van Staaden MJ, Kaufman LS, Liem KF. 1997 Microhabitat use, trophic patterns, and the evolution of brain structure in African cichlids. *Brain Behav. Evol.* **50**, 167–182. (doi:10.1159/000113330)
- Kotrschal K, Van Staaden MJ, Huber R. 1998 Fish brains: evolution and environmental relationships. *Rev. Fish Biol. Fisheries* **8**, 373–408. (doi:10.1023/A:1008839605380)
- Aristide L, Dos Reis SF, Machado AC, Lima I, Lopes RT, Perez SI. 2015 Encephalization and diversification of the cranial base in platyrrhine primates. *J. Hum. Evol.* **81**, 29–40. (doi:10.1016/j.jhevol.2015.02.003)
- Sol D, Price TD. 2008 Brain size and the diversification of body size in birds. *Am. Nat.* **172**, 170–177. (doi:10.1086/589461)
- Martin RD. 1990 *Primate origins and evolution: a phylogenetic reconstruction*. Princeton, NJ: Princeton University Press.
- Shultz S, Dunbar R. 2010 Species differences in executive function correlate with hippocampus volume and neocortex ratio across nonhuman primates. *J. Comp. Psychol.* **124**, 252. (doi:10.1037/a0018894)
- Passingham RE, Wise SP. 2012 *The neurobiology of the prefrontal cortex: anatomy, evolution, and the origin of insight*. Oxford, UK: Oxford University Press.
- Healy SD, Rowe C. 2007 A critique of comparative studies of brain size. *Proc. R. Soc. B* **274**, 453–464. (doi:10.1098/rspb.2006.3748)
- Weisbecker V, Blomberg S, Goldizen AW, Brown M, Fisher D. 2015 The evolution of relative brain size in marsupials is energetically constrained but not driven by behavioral complexity. *Brain Behav. Evol.* **85**, 125–135. (doi:10.1159/000377666)
- Byrne RW, Corp N. 2004 Neocortex size predicts deception rate in primates. *Proc. R. Soc. Lond. B* **271**, 1693. (doi:10.1098/rspb.2004.2780)
- Kudo H, Dunbar R. 2001 Neocortex size and social network size in primates. *Anim. Behav.* **62**, 711–722. (doi:10.1006/anbe.2001.1808)
- Pawłowski B, Lowen C, Dunbar R. 1998 Neocortex size, social skills and mating success in primates. *Behaviour* **135**, 357–368. (doi:10.1163/156853998793066285)
- Charvet CJ, Striedter GF, Finlay BL. 2011 Evo-devo and brain scaling: candidate developmental mechanisms for variation and constancy in vertebrate brain evolution. *Brain Behav. Evol.* **78**, 248–257. (doi:10.1159/000329851)
- Jerison HJ. 1973 *Evolution of the brain and intelligence*. New York, NY: Academic Press.
- Jerison HJ. 1990 Fossil evidence on the evolution of the neocortex. In *Comparative structure and evolution of cerebral cortex, Part I* (eds E Jones, A Peters), pp. 285–309. New York, NY: Springer US.
- Northcutt RG, Kaas JH. 1995 The emergence and evolution of mammalian neocortex. *Trends Neurosci.* **18**, 373–379. (doi:10.1016/0166-2236(95)93932-N)
- Medina L, Reiner A. 2000 Do birds possess homologues of mammalian primary visual,



- somatosensory and motor cortices? *Trends Neurosci.* **23**, 1–12. (doi:10.1016/S0166-2236(01)486-1)
35. Molnár Z, Tavare A, Cheung AFP. 2007 3.02—the origin of neocortex: lessons from comparative embryology. In *Evolution of nervous systems* (ed. JH Kaas), pp. 13–26. Oxford, UK: Academic Press.
36. Kaas JH. 2011 Neocortex in early mammals and its subsequent variations. *Ann. NY Acad. Sci.* **1225**, 28–36. (doi:10.1111/j.1749-6632.2011.05981.x)
37. Anthony R. 1938 Anatomie comparée du cerveau (Doin, Paris 1928)—Essai de recherche d'une expression anatomique approximative du degré d'organisation cérébrale, autre que le poids de l'encéphale comparé au poids du corps. *Bulletins et Mémoires de la Société d'anthropologie de Paris* **9**, 17–67. (doi:10.3406/bmsap.1938.9338)
38. Wirz K. 1950 Studien über die cerebralisation: Zur quantitativen bestimmung der rangordnung bei säugetieren. *Acta Anatomica* **9**, 134–196. (doi:10.1159/000140431)
39. Sawaguchi T, Kudo H. 1990 Neocortical development and social structure in primates. *Primates* **31**, 283–289. (doi:10.1007/BF02380949)
40. Changizi MA. 2001 Principles underlying mammalian neocortical scaling. *Biol. Cybern.* **84**, 207–215. (doi:10.1007/s004220000205)
41. Frahm HD, Stephan H, Stephan M. 1982 Comparison of brain structure volumes in Insectivora and Primates. I. Neocortex. *J. Hirnforschung* **23**, 375–389.
42. Changizi MA, Shimojo S. 2005 Parcellation and area-area connectivity as a function of neocortex size. *Brain Behav. Evol.* **66**, 88–98. (doi:10.1159/000085942)
43. Kaas JH. 1989 The evolution of complex sensory systems in mammals. *J. Exp. Biol.* **146**, 165–176.
44. Dunbar RIM. 1992 Neocortex size as a constraint on group-size in primates. *J. Hum. Evol.* **22**, 469–493. (doi:10.1016/0047-2484(92)90081-J)
45. Joffe TH, Dunbar R. 1997 Visual and socio-cognitive information processing in primate brain evolution. *Proc. R. Soc. Lond. B* **264**, 1303–1307. (doi:10.1098/rspb.1997.0180)
46. Connor R, Mann J, Tyack P, Whitehead H. 1998 Reply from Quantifying brain-behavior relations in cetaceans and primates. *Trends Ecol. Evol.* **13**, 408. (doi:10.1016/S0169-5347(98)01459-1)
47. Kaas JH. 2012 The evolution of neocortex in primates. *Progr. Brain Res.* **195**, 91–102. (doi:10.1016/B978-0-444-53860-4.00005-2)
48. Kaas JH. 1989 Why does the brain have so many visual areas? *J. Cogn. Neurosci.* **1**, 121–135. (doi:10.1162/jocn.1989.1.2.121)
49. Kaas JH. 2013 The evolution of brains from early mammals to humans. *Wiley Interdiscip. Rev. Cogn. Sci.* **4**, 33–45. (doi:10.1002/wcs.1206)
50. DeFelipe J. 2011 The evolution of the brain, the human nature of cortical circuits and intellectual creativity. *Front. Neuroanat.* **5**, 29. (doi:10.3389/fnana.2011.00029)
51. Letinic K, Zoncu R, Rakic P. 2002 Origin of GABAergic neurons in the human neocortex. *Nature* **417**, 645–649. (doi:10.1038/nature00779)
52. Aiello LC, Dunbar RIM. 1993 Neocortex size, group-size, and the evolution of language. *Curr. Anthropol.* **34**, 184–193. (doi:10.1086/204160)
53. Allison T, Ginter H, McCarthy G, Nobre AC, Puce A, Luby M, Spencer DD. 1994 Face recognition in human extrastriate cortex. *J. Neurophysiol.* **71**, 821–825.
54. Nestor A, Plaut DC, Behrmann M. 2011 Unraveling the distributed neural code of facial identity through spatiotemporal pattern analysis. *Proc. Natl Acad. Sci. USA* **108**, 9998–10 003. (doi:10.1073/pnas.1102433108)
55. Poucet B, Lenck-Santim PP, Paz-Villagran V, Save E. 2003 Place cells, neocortex and spatial navigation: a short review. *J. Physiol. Paris* **97**, 537–546. (doi:10.1016/j.jphysparis.2004.01.011)
56. Miller VM, Best PJ. 1980 Spatial correlates of hippocampal unit-activity are altered by lesions of the fornix and entorhinal cortex. *Brain Res.* **194**, 311–323. (doi:10.1016/0006-8993(80)91214-7)
57. Platek SM, Keenan JP, Gallup Jr GG, Mohamed FB. 2004 Where am I? The neurological correlates of self and other. *Cogn. Brain Res.* **19**, 114–122. (doi:10.1016/j.cogbrainres.2003.11.014)
58. Platek SM, Wathne K, Tierney NG, Thomson JW. 2008 Neural correlates of self-face recognition: an effect-location meta-analysis. *Brain Res.* **1232**, 173–184. (doi:10.1016/j.brainres.2008.07.010)
59. Sugiura M, Watanabe J, Maeda Y, Matsue Y, Fukuda H, Kawashima R. 2005 Cortical mechanisms of visual self-recognition. *Neuroimage* **24**, 143–149. (doi:10.1016/j.neuroimage.2004.07.063)
60. Crick F, Koch C. 1990 *Towards a neurobiological theory of consciousness*. Seminars in the neurosciences, pp. 263–275. Philadelphia, PA: Saunders Scientific Publications.
61. Eccles J. 1994 The evolution of consciousness. In *How the self controls its brain*, pp. 113–124. Berlin, Germany: Springer.
62. Gilbert P, Price J, Allan S. 1995 Social comparison, social attractiveness and evolution: how might they be related? *New Ideas Psychol.* **13**, 149–165. (doi:10.1016/0732-118X(00)02-X)
63. Grober E, Ausubel R, Sliwinski M, Gordon B. 1992 Skill learning and repetition priming in Alzheimer's disease. *Neuropsychologia* **30**, 849–858. (doi:10.1016/0028-3932(92)90030-P)
64. Greig LC, Woodworth MB, Galazo MJ, Padmanabhan H, Macklis JD. 2013 Molecular logic of neocortical projection neuron specification, development and diversity. *Nat. Rev. Neurosci.* **14**, 755–769. (doi:10.1038/nrn3586)
65. Shaw P *et al.* 2008 Neurodevelopmental trajectories of the human cerebral cortex. *J. Neurosci.* **28**, 3586–3594. (doi:10.1523/JNEUROSCI.5309-07.2008)
66. de Sousa AA, Proulx MJ. 2014 What can volumes reveal about human brain evolution? A framework for bridging behavioral, histometric, and volumetric perspectives. *Front. Neuroanat.* **8**, 51. (doi:10.3389/fnana.2014.00051)
67. Hawrylycz MJ *et al.* 2012 An anatomically comprehensive atlas of the adult human brain transcriptome. *Nature* **489**, 391–399. (doi:10.1038/nature11405)
68. Dorus S, Vallender EJ, Evans PD, Anderson JR, Gilbert SL, Mahowald M, Wyckoff GJ, Malcom CM, Lahn BT. 2004 Accelerated evolution of nervous system genes in the origin of *Homo sapiens*. *Cell* **119**, 1027–1040. (doi:10.1016/j.cell.2004.11.040)
69. Shi P, Bakewell MA, Zhang J. 2006 Did brain-specific genes evolve faster in humans than in chimpanzees? *Trends Genet.* **22**, 608–613. (doi:10.1016/j.tig.2006.09.001)
70. Kosiol C, Vinař T, da Fonseca RR, Hubisz MJ, Bustamante CD, Nielsen R, Siepel A. 2008 Patterns of positive selection in six mammalian genomes. *PLoS Genet.* **4**, e1000144. (doi:10.1371/journal.pgen.1000144)
71. Gutierrez H, Castillo A, Monzon J, Urrutia AO. 2011 Protein amino acid composition: a genomic signature of encephalization in mammals. *PLoS ONE* **6**, e27261. (doi:10.1371/journal.pone.0027261)
72. Lynch M, Conery JS. 2000 The evolutionary fate and consequences of duplicate genes. *Science* **290**, 1151–1155. (doi:10.1126/science.290.5494.1151)
73. Krylov DM, Wolf YI, Rogozin IB, Koonin EV. 2003 Gene loss, protein sequence divergence, gene dispensability, expression level, and interactivity are correlated in eukaryotic evolution. *Genome Res.* **13**, 2229–2235. (doi:10.1101/gr.1589103)
74. Demuth JP, De Bie T, Stajich JE, Cristianini N, Hahn MW. 2006 The evolution of mammalian gene families. *PLoS ONE* **1**, e85. (doi:10.1371/journal.pone.0000085)
75. Hahn MW, Han MV, Han SG. 2007 Gene family evolution across 12 *Drosophila* genomes. *PLoS Genet.* **3**, e197. (doi:10.1371/journal.pgen.0030197)
76. Hahn MW, De Bie T, Stajich JE, Nguyen C, Cristianini N. 2005 Estimating the tempo and mode of gene family evolution from comparative genomic data. *Genome Res.* **15**, 1153–1160. (doi:10.1101/gr.3567505)
77. Hoover KC. 2013 Evolution of olfactory receptors. *Methods Mol. Biol.* **1003**, 241–249. (doi:10.1007/978-1-62703-377-0\_18)
78. Malnic B, Hirono J, Sato T, Buck LB. 1999 Combinatorial receptor codes for odors. *Cell* **96**, 713–723. (doi:10.1016/S0092-8674(00)80581-4)
79. Kajiji K, Inaki K, Tanaka M, Haga T, Kataoka H, Touhara K. 2001 Molecular bases of odor discrimination: reconstitution of olfactory receptors that recognize overlapping sets of odorants. *J. Neurosci.* **21**, 6018–6025.
80. Niimura Y, Matsui A, Touhara K. 2014 Extreme expansion of the olfactory receptor gene repertoire in African elephants and evolutionary dynamics of orthologous gene groups in 13 placental mammals. *Genome Res.* **24**, 1485–1496. (doi:10.1101/gr.169532.113)
81. Castillo-Morales A, Monzon-Sandoval J, Urrutia AO, Gutierrez H. 2014 Increased brain size in mammals is associated with size variations in gene families with cell signalling, chemotaxis and immune-related functions. *Proc. R. Soc. B* **281**, 20132428. (doi:10.1098/rspb.2013.2428)



82. Cunningham F *et al.* 2015 Ensembl 2015. *Nucleic Acids Res.* **43**(Database issue), D662–D669. (doi:10.1093/nar/gku1010)
83. Gonzalez-Lagos C, Sol D, Reader SM. 2010 Large-brained mammals live longer. *J. Evol. Biol.* **23**, 1064–1074. (doi:10.1111/j.1420-9101.2010.01976.x)
84. Pirlot P, Jiao SS. 1985 Quantitative morphology of the panda brain in comparison with the brains of the raccoon and the bear. *J. Hirnforschung* **26**, 17–22.
85. Stephan H, Frahm H, Baron G. 1981 New and revised data on volumes of brain structures in insectivores and primates. *Folia Primatol. Int. J. Primatol.* **35**, 1–29. (doi:10.1159/000155963)
86. Schleifenbaum C. 1973 Untersuchungen zur postnatalen Ontogenese des Gehirns von Großspudeln und Wölfen. *Z. Anat. Entwickl. Gesch.* **141**, 179–205. (doi:10.1007/BF00519885)
87. Kruska DCT. 2014 Comparative quantitative investigations on brains of wild cavies (*Cavia aperea*) and guinea pigs (*Cavia aperea f. porcellus*). A contribution to size changes of CNS structures due to domestication. *Mamm. Biol.* **79**, 230–239. (doi:10.1016/j.mambio.2013.12.005)
88. Hakeem AY, Hof PR, Sherwood CC, Switzer RC, Rasmussen LEL, Allman JM. 2005 Brain of the African elephant (*Loxodonta africana*): neuroanatomy from magnetic resonance images. *Anat. Rec. A Discov. Mol. Cell Evol. Biol.* **287A**, 1117–1127. (doi:10.1002/ar.a.20255)
89. Pirlot P. 1981 A quantitative approach to the marsupial brain in an eco-ethological perspective. *Revue canadienne de biologie/editee par l'Universite de Montreal* **40**, 229–250.
90. Beatty J, Laughlin RE. 2006 Genomic regulation of natural variation in cortical and noncortical brain volume. *BMC Neurosci.* **7**, 16. (doi:10.1186/1471-2202-7-16)
91. Roehrs M. 1986 Cephalization, telencephalization and neocorticalization within Mustelidae. *Zeitschrift für Zoologische Systematik und Evolutionsforschung* **24**, 157–166. (doi:10.1111/j.1439-0469.1986.tb00624.x)
92. Pirlot PNJ. 1978 *Volumetric analyses of monotreme brains*. Sydney, Australia: The Royal Zoological Society of New South Wales.
93. Ebinger P. 1974 A cytoarchitectonic volumetric comparison of brains in wild and domestic sheep. *Z. Anat. Entwicklungsgesch* **144**, 267–302. (doi:10.1007/BF00522811)
94. Zilles K, Rehkämper G. 1988 *The brain, with special reference to the telencephalon*. Oxford, UK: Oxford University Press.
95. Bush EC, Allman JM. 2004 The scaling of frontal cortex in primates and carnivores. *Proc. Natl Acad. Sci. USA* **101**, 3962–3966. (doi:10.1073/pnas.0305760101)
96. Baron G, Stephan H, Frahm HD. 1996 *Comparative Neurobiology in Chiroptera: brain characteristics in functional systems, ecoethological adaptation, adaptive radiation, and evolution*. Basel, Switzerland: Birkhäuser Verlag.
97. Kruska D, Rohrs M. 1974 Comparative–quantitative investigations on brains of feral pigs from the Galapagos Islands and of European domestic pigs. *Z. Anat. Entwicklungsgesch* **144**, 61–73. (doi:10.1007/BF00518633)
98. Tschudin AJ-PC. 1998 Relative neocortex size and its correlates in dolphins: comparisons with humans and implications for mental evolution. PhD thesis, University of KwaZulu-Natal.
99. Kruska D. 1980 Domestikationsbedingte Hirngrößenänderungen bei Säugetieren. *Z. Zool. Syst. Evol.* **18**, 161–195. (doi:10.1111/j.1439-0469.1980.tb00738.x)
100. Tacutu R, Craig T, Budovsky A, Wuttke D, Lehmann G, Taranukha D, Costa J, Fraifield VE, de Magalhaes JP. 2013 Human Ageing Genomic Resources: integrated databases and tools for the biology and genetics of ageing. *Nucleic Acids Res.* **41**(Database issue), D1027–D1033. (doi:10.1093/nar/gks1155)
101. Felsenstein J. 1985 Phylogenies and the comparative method. *Am. Nat.* **125**, 1–15. (doi:10.2307/2461605)
102. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
103. Kumar S, Hedges SB. 2011 TimeTree2: species divergence times on the iPhone. *Bioinformatics* **27**, 2023–2024. (doi:10.1093/bioinformatics/btr315)
104. Miller JA *et al.* 2014 Transcriptional landscape of the prenatal human brain. *Nature* **508**, 199–206. (doi:10.1038/nature13185)
105. Dunbar RI, Shultz S. 2007 Understanding primate brain evolution. *Phil. Trans. R. Soc. B* **362**, 649–658. (doi:10.1098/rstb.2006.2001)
106. Kaas JH, Gharbawie OA, Stepniwska I. 2013 Cortical networks for ethologically relevant behaviors in primates. *Am. J. Primatol.* **75**, 407–414. (doi:10.1002/ajp.22065)
107. Prieto AL, Weber JL, Lai C. 2000 Expression of the receptor protein-tyrosine kinases Tyro-3, Axl, and Mer in the developing rat central nervous system. *J. Comp. Neurol.* **425**, 295–314. (doi:10.1002/1096-9861(20000918)425:2<295::AID-CNE11>3.0.CO;2-G)
108. Hafizi S, Dam B. 2006 Signalling and functional diversity within the Axl subfamily of receptor tyrosine kinases. *Cytokine Growth Factor Rev.* **17**, 295–304. (doi:10.1016/j.cytogfr.2006.04.004)
109. Allen MP, Linseman DA, Udo H, Xu M, Schaack JB, Varnum B, Kandel ER, Heidenreich KA, Wierman ME. 2002 Novel mechanism for gonadotropin-releasing hormone neuronal migration involving Gas6/Ark signaling to p38 mitogen-activated protein kinase. *Mol. Cell Biol.* **22**, 599–613. (doi:10.1128/MCB.22.2.599-613.2002)
110. Wada K, Arita M, Nakajima A, Katayama K, Kudo C, Kamisaki Y, Serhan CN. 2006 Leukotriene B4 and lipoxin A4 are regulatory signals for neural stem cell proliferation and differentiation. *FASEB J.* **20**, 1785–1792. (doi:10.1096/fj.06-5809com)
111. Gutierrez H, Davies AM. 2011 Regulation of neural process growth, elaboration and structural plasticity by NF-kappaB. *Trends Neurosci.* **34**, 316–325. (doi:10.1016/j.tins.2011.03.001)
112. Monzon-Sandoval J, Castillo-Morales A, Crampton S, McKelvey L, Nolan A, O'Keeffe G, Gutierrez H. 2015 Modular and coordinated expression of immune system regulatory and signaling components in the developing and adult nervous system. *Front. Cell. Neurosci.* **9**, 337. (doi:10.3389/fncel.2015.00337)
113. Keeney JG, Dumas L, Sikela JM. 2014 The case for DUF1220 domain dosage as a primary contributor to anthropoid brain expansion. *Front. Hum. Neurosci.* **8**, 427. (doi:10.3389/fnhum.2014.00427)
114. Keeney JG, Davis JM, Siegenthaler J, Post MD, Nielsen BS, Hopkins WD, Sikela JM. 2015 DUF1220 protein domains drive proliferation in human neural stem cells and are associated with increased cortical volume in anthropoid primates. *Brain Struct. Funct.* **220**, 3053–3060. (doi:10.1007/s00429-014-0814-9)
115. Dumas LJ *et al.* 2012 DUF1220-domain copy number implicated in human brain-size pathology and evolution. *Am. J. Hum. Genet.* **91**, 444–454. (doi:10.1016/j.ajhg.2012.07.016)
116. Dumas L, Sikela JM. 2009 DUF1220 domains, cognitive disease, and human brain evolution. *Cold Spring Harb. Symp. Quant. Biol.* **74**, 375–382. (doi:10.1101/sqb.2009.74.025)
117. Lui JH, Hansen DV, Kriegstein AR. 2011 Development and evolution of the human neocortex. *Cell* **146**, 18–36. (doi:10.1016/j.cell.2011.06.030)
118. Maaser K, Borlak J. 2008 A genome-wide expression analysis identifies a network of EpCAM-induced cell cycle regulators. *Br. J. Cancer* **99**, 1635–1643. (doi:10.1038/sj.bjc.6604725)
119. Gaiser MR, Lammermann T, Feng X, Igyarto BZ, Kaplan DH, Tessarollo L, Germain RN, Udey MC. 2012 Cancer-associated epithelial cell adhesion molecule (EpCAM; CD326) enables epidermal Langerhans cell motility and migration *in vivo*. *Proc. Natl Acad. Sci. USA* **109**, E889–E897. (doi:10.1073/pnas.1117674109)
120. Gastwirt RF, McAndrew CW, Donoghue DJ. 2007 Speedy/RINGO regulation of CDKs in cell cycle, checkpoint activation and apoptosis. *Cell Cycle* **6**, 1188–1193. (doi:10.4161/cc.6.10.4252)
121. Lenormand JL, Dellinger RW, Knudsen KE, Subramani S, Donoghue DJ. 1999 Speedy: a novel cell cycle regulator of the G2/M transition. *EMBO J.* **18**, 1869–1877. (doi:10.1093/emboj/18.7.1869)
122. Chauhan S, Zheng X, Tan YY, Tay BH, Lim S, Venkatesh B, Kaldis P. 2012 Evolution of the Cdk-activator Speedy/RINGO in vertebrates. *Cell. Mol. Life Sci.* **69**, 3835–3850. (doi:10.1007/s00018-012-1050-1)
123. Zhang Y *et al.* 2013 Functional genomic screen of human stem cell differentiation reveals pathways involved in neurodevelopment and neurodegeneration. *Proc. Natl Acad. Sci. USA* **110**, 12 361–12 366. (doi:10.1073/pnas.1309725110)
124. Andreasen NC. 2010 The lifetime trajectory of schizophrenia and the concept of neurodevelopment. *Dialogues Clin. Neurosci.* **12**, 409–415.